

Synthesis and revision of the relative configuration of fudecalone

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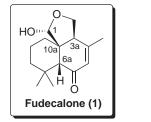
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Abstract—We synthesized the proposed structure of fudecalone, whose NMR spectral data were not identical with those of the natural compound. As our synthetic fudecalone was a conformational isomer of the reported structure, we first investigated conformational isomerization. However, all our attempts resulted in failure, and we then changed the strategy to the synthesis of diastereomers. Finally, we found the *trans*-octalone derivative showed the identical NMR spectrum with that of natural fudecalone, and the relative configuration was determined to be $1S^*$, $3aS^*$, $6aS^*$, $10aS^*$. © 2001 Elsevier Science Ltd. All rights reserved.

Fudecalone (1) was isolated by \overline{O} mura et al. in 1995 from a culture broth of *Penicillium* sp. FO-2030 as an anticoccidial sesquiterpene. It completely inhibited schizont formation of monensin-resistant *Eimeria tenella* at concentrations of more than 16 μ M. The structure was elucidated mainly by NMR experiments and the conformation was reported to be 1a as illustrated in Fig. 1.

As reported previously, we started our synthesis from the iodide (2) and the known phthalide (3), and obtained 5 via a keto-lactone (4) in six steps.² However, the ¹H and ¹³C NMR spectral data of 5 were not identical with those of reported data and an analysis of its NOESY spectrum revealed the conformation of 5 to be 1b, which was a conformational isomer of 1a (Scheme 1).

According to MM3 calculation, **1b** was 2.1 kcal/mol more stable than **1a** (Fig. 2). So, we thought it would be



HO Ta

HO extra-high energy barrier?

Figure 1.

Figure 2.

Scheme 1.

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Scheme 2. (a) TBSOTf, 2,6-lut. CH₂Cl₂, 0°C (92%); (b) H₂, Pd–C, EtOAc, rt (82%, diastereo ratio was 4:1); (c) TBAF, THF, rt (83%); (d) DIBAL, CH₂Cl₂, -78°C; (e) *p*-TsOH, MeOH, rt (49% in two steps); (f) Dess–Martin periodinane, CH₂Cl₂, rt (83%); (g) LDA, THF; PhSeCl, HMPA, -78°C (82%); (h) *m*CPBA, CH₂Cl₂, 0°C (53%).

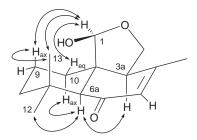
Scheme 3. (a) H_2 , Pd–C, EtOH, rt (88%); (b) aq. LiOH, THF, reflux; (c) PPTS, benzene, reflux (69% in two steps, 92% based on recovered 12); (d) LDA, THF; TMSCl, Et₃N, -78°C (91%); (e) NBS, THF, rt (93%); (f) DBU, THF, reflux (93%); (g) DIBAL, tol. -78°C; (h) MnO₂, CH₂Cl₂, rt (32% in two steps).

interesting to know if fudecalone could exist as an unstable conformation because of the extra-high energy barrier between 1a and 1b, and we started an investigation of the conformational isomerization from 1b to 1a.

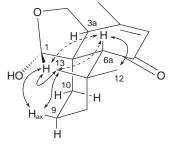
We adopted intermediate (4) which was dienylsilylated with TBSOTf followed by hydrogenation to give silylether (7) (Scheme 2). This hydrogenation to exoolefin of $\boldsymbol{6}$ occurred predominantly from the α face $(\alpha:\beta=4:1)$. Treatment of 7 with TBAF afforded a ketone (8) in 50% yield over three steps. We expected that the conformation of 8 would change from 1b type to 1a type so that the β-methyl group would be stabilized in a more stable equatorial orientation. However, very strong NOE between H-4 and H₃-12 revealed that the B-ring was in the boat-form, and thus 6 still kept the 1b-type conformation. When 8 was reduced to 9 with 4 equiv. of DIBAL, the conformation changed to 1a type for the first time. As 9 was unstable, we converted it into a methylacetal with p-TsOH in MeOH (49% yield from 8), which was oxidized with Dess-Martin periodinane³ followed by selenylation to give 10. The NOE between H-1 and H_3 -13 revealed that 10 retained the same conformation. But when the selenide was eliminated by oxidizing with mCPBA, the conformation of the product reverted to the previous 1b type, and 11 was obtained in 53% yield. The conformation of 11 was confirmed by NOE experiments and X-ray analysis.

From these results, it was clarified that the energy barrier between 1a and 1b was not so high and that this compound could not exist as 1a but only as 1b. We concluded therefore that the proposed relative stereochemistry was incorrect and decided to synthesize a *trans*-fused octalone, 1c.

We started a synthesis of 1c from the same intermediate (4), which was hydrogenated with Pd-C to afford 12 as an almost single isomer (Scheme 3). This direct hydrogenation made the methyl group of 12 α -oriented, because the α -face of the B-ring is sterically protected



Synthetic Fudecalone 1c



Reported Fudecalone 1a

Figure 3.

by the 7-methyl group. In the presence of the lactone ring, cis-fused decalone 12 could not be isomerized to trans-fused decalone 15 by using an organic base, DBU, LDA or LHMDS. However, when treated with ag. LiOH, the lactone was saponified first. Then the conformation of hydroxycarboxylic acid changed to 13 and the equilibrium favored 14, and then 15 was obtained after the relactonization with PPTS under reflux in benzene for 4 h in 69% yield over two steps. Thus, the lactone-opening was essential for this isomerization. On the other hand, 15 completely isomerized to the more stable 12 by heating with PPTS for a longer period. The enolate derived from 15 by LDA treatment was trapped with TMSCl followed by bromination, dehydrobromination afforded an enone (16) in 79% yield over three steps, and the stereochemistry of 16 was confirmed by X-ray analysis. The unsaturated ketone and the lactone carbonyls of 16 were reduced with 4.4 equiv. of DIBAL, and the resultant allylic alcohol was reoxdized with MnO₂ to furnish 1c in 32% yield over two steps. ¹H and ¹³C NMR spectral data of 1c showed complete accordance with that of natural fudecalone except the NOE data.4

The NOESY spectrum of our 1c showed slight differences from the reported data, as illustrated in Fig. 3. NOEs of 1c between H-6a and each of H-3a, H- 10_{ax} and H₃-12 indicated that C-6a had the S^* configuration, and NOEs between H-1 and each of H- 9_{ax} , H- 10_{eq} and H₃-13 indicated that C-1 also had the S^* configuration. On the other hand, NOEs between H-6a and each of H-1 and H₃-13 of 1 indicated by dashed arrows, which were the decisive factor to determine fudecalone to be 1a, were not observed.

In conclusion, our investigation of the conformational isomerization from **1b** to **1a** via selenide (**10**) proved fruitless. However, by utilizing the intermediate of the previous synthesis of **1b**, we completed the synthesis of *trans*-fused octalone **1c** as a racemate and found that it showed the identical NMR spectral data with those of

the natural fudecalone. By further NOESY experiments we were able to determine the relative configuration of fudecalone to be $1S^*$, $3aS^*$, $6aS^*$ and $10aS^*$.

Enantioselective synthesis and the determination of the absolute configuration of natural fudecalone are in progress and will be reported in a full account.

Acknowledgements

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References

- Tabata, N.; Tomoda, H.; Masuda, R.; Iwai, Y.; Ōmura, S. J. Antibiot. 1995, 48, 53–58.
- Watanabe, H.; Furuuchi, T.; Yamaguchi, T.; Kitahara, T. Org. Lett. 1999, 1, 1079–1080.
- (a) Dess, D. B.; Martin, J. C. J. Org. Chem. 1983, 48, 4155–4156; (b) Dess, D. B.; Martin, J. C. J. Am. Chem. Soc. 1991, 113, 7277–7287; (c) Ireland, R. E.; Lui, L. J. Org. Chem. 1993, 58, 2899; (d) Meyer, S. D.; Schreiber, S. L. J. Org. Chem. 1994, 59, 7549–7552.
- 4. ¹H NMR (500 MHz, in CDCl₃) δ 1.16 (3H, s), 1.24 (3H, s), 1.25 (1H, dt, J=4.0, 13.5 Hz), 1.26 (1H, dt, J=4.0, 13.5 Hz), 1.44 (1H, m), 1.53 (1H, quint. t, J=3.5, 14.0 Hz), 1.63 (1H, tq, J=3.5, 14.0 Hz), 1.82 (3H, s), 1.86 (1H, m), 2.38 (1H, s), 2.46 (1H, d, J=3.5 Hz), 2.61 (1H, dd, J=4.0, 9.0 Hz), 3.96 (1H, dd, J=4.0, 9.0 Hz), 4.24 (1H, t, J=9.0 Hz), 5.47 (1H, d, J=3.5 Hz), 5.82 (1H, s); ¹³C NMR (125 MHz, in CDCl₃) δ 19.62, 21.41, 21.78, 32.20, 32.60, 37.59, 43.07, 51.09, 51.97, 56.53, 68.41, 100.87, 128.58, 153.11, 197.74; mp 186–196°C.